XANES Spectroscopy for Monitoring Intermediate Reaction States of Cl⁻-Depleted Mn Cluster in Photosynthetic Water Oxidation Enzyme

Taka-aki Ono,*.[†] Takumi Noguchi,[†] Yorinao Inoue,[†] Masami Kusunoki,[‡] Hirotaka Yamaguchi,[§] and Hiroyuki Oyanagi§

Solar Energy Research Group, The Institute of Physical and Chemical Research (RIKEN) Wako, Saitama 351-01, Japan School of Science and Technology, Meiji University Kawasaki, Kanagawa 214, Japan Electromechanical Laboratory Tsukuba, Ibaraki 305, Japan

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The structural and electronic states of the Mn cluster in Cl⁻depleted photosynthetic water oxidation enzyme (WOE) were directly monitored by means of X-ray absorption near-edge structure (XANES) spectroscopy. Changes in XANES spectra of the Mn cluster in each intermediate reaction state induced by successive flash illumination provide direct evidence that a Mn ion is oxidized from Mn(III) to Mn(IV) after one flash, but further oxidation steps are interrupted in the absence of Cl⁻. The normal Mn K-edge energy, however, suggests that the ligation structure of the Mn cluster does not change much after depletion of Cl⁻.

The photosynthetic water oxidation is catalyzed by the tetranuclear Mn cluster¹ located in the photosystem II (PSII) core complex and proceeds through four stable intermediate reaction states of WOE, denoted as S_i (i = 0-3), by use of the energy of four photons.² Our previous XANES experiments with O₂-evolving WOE have revealed that the energy of Mn K-edge shows a period-four oscillation depending on each intermediate S-state induced by a short laser \overline{flash} .^{3,4} Cl⁻ is an indispensable inorganic cofactor for photosynthetic water oxidation and also functions in structural stabilization of the Mn cluster,⁵ but the mode of Cl⁻ action in the water oxidation has remained largely unknown. One slowly exchanging Cl⁻ that is involved in the O_2 evolution exists in PSII and is released almost completely from its site by a NaCl wash.⁶ In the absence of Cl⁻, a limited number of oxidizing equivalents for water oxidation are accumulated on the donor side of PSII,⁵ although their chemical identity is still a matter of debate.⁷ Under these circumstances, it is necessary to probe directly the Mn cluster in order to characterize the redox reactions occurring in the Cl⁻depleted WOE. In this report, we adopted XANES spectroscopy to directly probe the electronic state of the Mn cluster. PSII

(1) Sauer, K.; Yachanda, V. K.; Britt, R. D.; Klein, M. P. In Manganese Redox Enzyme; Pecoraro, V. L., Ed.; VHC: New York, 1992; pp 105-

(6) Lindberg, K.; Wydrzynski, T.; Vänngard, T.; Andréasson, L.-E. FEBS Lett. 1990, 264, 153–155.

(7) Boussac, A.; Rutherford, A. W. J. Biol. Chem. 1994, 269, 12462-12467.

membranes capable of O_2 evolution⁸ were relaxed for 6 h in darkness and then subjected to a 2 M NaCl wash to facilitate the release of Cl⁻ from WOE.^{6,9} For Cl⁻ depletion, the treated membranes were then washed three times with a Cl⁻-free medium containing 0.4 M sucrose and 20 mM Mes-NaOH (pH 6.5) and suspended in the same medium supplemented with 20 mM Ca(OH)₂-Mes (pH 6.5). O_2 evolution activities of the control, Cl⁻-depleted, and Cl⁻-repleted membranes were 610, 90, and 540 μ mol of O₂/mg of chlorophyll/h, respectively. The resulting Cl⁻-depleted membranes were illuminated either with laser flashes or with continuous light and then immediately cooled to 77 K in liquid N2.4,5.10 XANES spectra were measured by fluorescence detection mode at the Photon Factory of the National Laboratory for High Energy Physics beam-line 4B.^{4,5,10} TL was measured as described previously.¹¹

K-edge XANES spectra of Cl⁻-depleted WOE after 0-5 flashes and continuous illumination were measured (Figure 1A), and their edge energies were compared in an expanded view in the K-edge region (Figure 1B). The half-height energy in darkadapted membranes was located at 6552.1 eV, which was almost the same value as that reported in the normal S_1 state with the same experimental setup.⁴ This indicates that the Mn cluster is retained intact after Cl⁻ depletion since even slight damage to the cluster is known to cause a downward shift of the K-edge energy due to the formation of Mn^{2+} . The structural intactness of the Mn cluster was also indicated by TL measurements: the Cl⁻-depleted membranes showed the TL band with an elevated peak temperature,¹¹ but the TL band due to the normal S₂ state was restored concomitant with a normal period-four flash pattern when sufficient Cl⁻ was added (data not shown). The halfheight energies of the spectra were determined to be 6552.8, 6552.9, 6552.7, 6552.5, and 6552.6 eV after 1, 2, 3, 4, and 5 flashes, respectively, and 6552.9 eV after continuous illumination

Figure 2 shows the flash number dependence of the halfheight energy of the Mn K-edge in Cl⁻-depleted WOE, where the flash pattern obtained in the nondepleted control WOE⁴ previously measured in the presence of sufficient Cl⁻ is shown as a reference. The K-edge energy shifted upward by 0.7 eV after 1 flash illumination of the Cl⁻-depleted WOE. This change was comparable to that induced after 1 flash in the control WOE, indicating oxidation of Mn(III) to Mn(IV).¹² This is the first direct evidence that the Mn cluster is oxidized in Cl⁻-depleted WOE. The K-edge energy did not undergo significant change after subsequent flashes in contrast to the control membranes, although a faint overlapping oscillation was apparent. After continuous illumination, the K-edge energy did not change much, compared with that after 1 flash. The residual oscillation exhibited a quadruple pattern as observed in the control WOE. Since the Cl⁻-depleted WOE showed about 15% of the O₂ evolution activity of the control, it can be reasonably assumed that the nondepleted WOE was responsible for this residual oscillation. The closed circles in Figure 2 represent the simulated pattern obtained by assuming an upshift of the energy after 1 flash but no oscillatory behavior after subsequent flashes in 85% WOE and the period four oscillation in 15% WOE. The simulation is in good agreement with the experimentally obtained pattern, indicating that the oscillation of the K-edge energy was interrupted in the Cl⁻-depleted WOE after 1 flash.

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^{*} To whom correspondence should be addressed. † The Institute of Physical and Chemical Research (RIKEN).

[‡] Meiji University.

[§] Electromechanical Laboratory

^{(2) (}a) Joliot, P.; Barbieri, G.; Chaud, R. Photochem. Photobiol. 1969, (2) (a) Johol, F.; Barbieri, G.; Chadd, R. Photochem. Photobiol. 1969, 10, 309-329. (b) Kok, B.; Forbush, B.; McGloin, M. Photochem. Photobiol. 1970, 11, 457-475.
(3) Ono, T.; Noguchi, T.; Inoue, Y.; Kusunoki, M.; Matsushita, T.; Oyanagi, H. Science 1992, 258, 1335-1337.
(4) Ono, T.; Noguchi, T.; Inoue, Y.; Kusunoki, M.; Yamaguchi, H.; Oyanagi, H. Biochem. Soc. Trans. 1994, 22, 331-335.
(5) For recent reviews, see: (a) Homann, P. H. J. Bioenerg. Biomembr. 1987 19 105-123. (b) Coleman J. W.; Goviniee Photosynth Res. 1987.

^{1987, 19, 105-123. (}b) Coleman, J. W.; Govindjee. Photosynth. Res. 1987, 13, 199–223. (c) Debus, R. J. Biochim. Biophys. Acta 1992, 1102, 269–352.
 (d) Rutherford, A. W.; Zimmermann, J.-L.; Boussac, A. In The Photosystem: Structure, Function and Molecular Biology; Barber, J., Ed.; Elsvier: Amsterdam, 1992; pp 179-229.

⁽⁸⁾ Berthold, D. A.; Babcock, G. T. and Yocum, C. F. FEBS Lett. 1981, 134, 231-234.

⁽⁹⁾ Miyao, M.; Murata, N. FEBS Lett. 1985, 180, 303-307.

⁽¹⁰⁾ Ono, T.; Noguchi, T.; Inoue, Y.; Kusunoki, M.; Yamaguchi, H.; Oyanagi, H. *FEBS Lett.* **1993**, *330*, 28–30.

⁽¹¹⁾ Homann, P. H.; Gleiter, H.; Ono, T.; Inoue, Y. Biochim. Biophys. Acta 1986, 850, 10-20

^{(12) (}a) Goodin, D. B.; Yachandra, V. K.; Britt, R. D.; Sauer, K.; Klein,
M. P. Biochim. Biophys. Acta 1984, 767, 209-216. (b) McDermott, A.
E.; Yachandra, V. K.; Guiles, R. D.; Cole, J. L.; Dexheimer, S. L.; Britt,
R. D.; Sauer, K.; Klein, M. P. Biochemistry 1988, 27, 4021-4031.

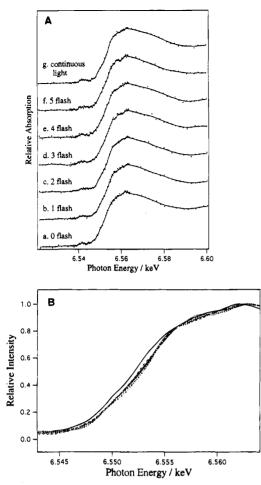


Figure 1. (A) XANES spectra of Cl⁻-depleted PSII membranes after a series of flashes and continuous illumination. Spectra a-f correspond to 0-5 flashes, and spectrum g is for continuous illumination at 0 °C for 1 min. (B) Effect of illumination on Mn K-edge. The membranes were illuminated with 0 (heavy solid line), 1 (light solid line), 2 (light short dash line), 3 (heavy short dash line), 4 (long dash line), and 5 (dotted line) flashes, and continuous illumination (dash-dot line). Fluorescence X-ray from the sample was detected with a solid state detector placed 10 mm from the sample. Samples were kept in darkness at 30 K during data collection. K-edge energies were calibrated using the pre-edge peak of KMnO4 at 6543.3 eV measured simultaneously by absorption mode. For each spectra, 7-9 scans were averaged.

In Cl⁻-depleted WOE, neither CW¹³ nor pulsed⁷ EPR detects any multiline EPR signal, but the multiline is developed by flash illumination followed by addition of Cl⁻ in the dark.¹³ The present result provides evidence that the Mn cluster is oxidized by 1 flash, as seen in the O₂-evolving WOE, but the resulting state (Cl⁻-depleted S_2 state) is silent to EPR. The fact that the K-edge energies for the S_1 and S_2 states are almost the same in the normal and Cl⁻-depleted WOE indicate that the EPR-silent S_2 state is not ascribed to a change in the oxidation state of the Mn ion but rather to a change in antiferromagnetic and ferromagnetic coupling between Mn ions.

It is interesting to note in this context that the K-edge energy after continuous illumination was quite similar to that after 1 flash. Continuous illumination of the Cl⁻-depleted WOE at 0 °C leads to the formation of a broad EPR signal at $g = 2.^{14.15}$ The g = 2 EPR signal was induced by illumination in inhibited WOEs and has been thought to arise from a free radical of

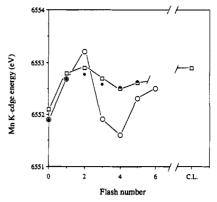


Figure 2. Flash-number-dependent changes of the half-height energy of the Mn K-edge after a series of flashes and continuous illumination (CL) in Cl⁻-depleted (\Box) and nontreated control (\bigcirc) PSII membranes. The simulated pattern (•) was obtained as described in the text. The K-edge energy was obtained from the smoothed spectra shown in Figure 1. An estimated error in determination of the K-edge energy was ± 0.1 eV.

histidine,¹⁶ tyrosine,¹⁷ or partially oxidized water¹⁸ interacting magnetically with the Mn cluster. It can, therefore, be concluded that no change in the K-edge energy is accompanied by the formation of the g = 2 EPR state in Cl⁻-depleted WOE. In contrast, the K-edge energy shifted upward by 0.9 eV concurrently with the formation of the g = 2 EPR state in Ca²⁺depleted WOE.¹⁰ In Ca²⁺-depleted WOE, the K-edge energy for the dark-adapted state (S1) was lowered by 1.5 eV compared to that of the normal S₁ state, suggesting a relatively large change in the ligation structure of the Mn cluster.¹⁰ Consistent with the marked downshift of the Mn K-edge energy, FT-IR difference spectroscopy reveals the breakage of the coordination bond by Ca^{2+} depletion between the redox active Mn ion and a certain carboxylate group that serves as a bridging ligand between the Mn cluster and Ca^{2+} in the normal S₁ state.¹⁹ The minimal effect of Cl⁻ depletion on the K-edge energy may, therefore, suggest that Cl⁻ depletion does not result in significant structural modification of the Mn cluster. It has been proposed that Cl⁻ functions as a direct ligand for the Mn cluster²⁰ and furthermore acts as a bridging ligand between Mn atoms to stabilize the structure of the Mn cluster.²¹ The present results, however, prefer the idea that Cl⁻ is not the first coordination ligand of the Mn cluster, although we cannot completely rule out the possibility of the direct ligation of Cl⁻ to the Mn cluster at present in a way that the lack of Cl⁻ does not cause a gross structural modification in the cluster.

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- (20) Yachandra, V. K.; Derose, V. J.; Latimer, M. J.; Mukeri, I.; Sauer, K.; Klein, M. P. Science 1993, 260, 675-682.
 (21) Sandusky, P. O.; Yocum, C. F. FEBS Lett. 1983, 162, 339-343.

⁽¹³⁾ Ono, T.; Zimmerman, J.-L.; Inoue, Y.; Rutherford, A. W. Biochim. Biophys. Acta. **1986**, 857, 1993–201. (14) Baumgarten, J.; Philo, J.; Dismukes, G. C. *Biochemistry* **1990**, 29,

^{10814 - 10822}

⁽¹⁵⁾ Boussac, A.; Setif, P.; Rutherford, A. W. Biochemistry 1992, 31, 1224-1234.

⁽¹⁶⁾ Boussac, A.; Zimmermann, J.-L.; Rutherford, A. W.; Lavergne, J. Nature 1990, 347, 303-306.

⁽¹⁷⁾ Hallahan, B. J.; Nugent, J. H.; Warden, J. T.; Evans, M. Biochemistry 1992, 31, 4562-4573.

⁽¹⁸⁾ Kusunoki, K. Plant Cell Physiol. 1993, 34 (Supplement), 55. (19) Noguchi, T.; Ono, T.; Inoue, Y. Biochim. Biophys. Acta. 1995, 1228, 189 - 200.